(Original Article)



Predicting the Severity of Alfalfa Root *rot* Disease Under Salinity Conditions

Khaled Hussein Arafat^{1*}; Mohammed H.A. Hassan² and Omar H.H. Mahmoud¹

¹Plant Pathology Department, Faculty of Agriculture, New Valley University. Egypt. ²Plant Pathology Department, Faculty of Agriculture, Assiut University, Egypt.

* Correspondence: drkhaledarafat@agr.nvu.edu.eg DOI: 10.21608/AJAS.2023.223841.1280 © Faculty of Agriculture, Assiut University

Abstract

Alfalfa roots are infected with several fungal pathogens under salinity conditions. The effect of two salts of NaCl and CaCO₃ at three concentrations was studied in vitro. Mycelial growth and disease severity (DS%) were estimated to determine the relationship between (DS%) and water salinity. The highest effect on mycelial growth was detected at the electrical conductivity (EC_w value) of 18.75 ds/m² which ranged from 8.80 and 9.44% with NaCl and CaCO₃, respectively. Furthermore, Exserohilum sp. was the most affected by water salinity. With the different levels of water salinity, the DS% of alfalfa root rot increased gradually with the increased water salinity level. The highest level of water salinity with NaCl and CaCO₃ EC_w value at 18.75 ds/m² ranged (42.76 and 43.53% severity for NaCl) and (44.04 and 44.42% severity for CaCO₃, respectively). Moreover, with water salinity NaCl and CaCO₃, Fusarium sp. was the highest (DS%) with NaCl. Alfalfa root rot prediction model based on information gathered from the interaction of pathogen, water salinity, disease severity, and root and shoot length. It was found the relation between DS% with fungi, salt type, salt concentration, root and shoot length in the multiple regression model ($r^2 = 88.83\%$).

Keywords: Alfalfa, Root rots, Water salinity, Prediction models

Introduction

Alfalfa (*Medicago sativa* L.), is a perennial herb native to Iran and Central Asia (Bingham *et al.*, 1975; Harrison *et al.*, 2002). Alfalfa is widely regarded as a superior feed for dairy and beef cattle (*Bos* spp.) because it is easily digested, has a high protein content and a high concentration of cell solutes. (Conrad and Klopfenstein, 1988).

The total area under cultivation in Egypt throw 2018/2019 was 73469 feddans and the total production was 3156757 tons, while in New Valley Governorate, the total cultivated area in 2018/2019 was 41612 feddans and the total production was 2307613 tons (CAPMAS, 2023).

Several biotic agents and abiotic agents highly stress effect on alfalfa yield. Root rot is one of the most damaging fungal diseases. Root rot pathogens are responsible for some of the most serious plant diseases in the world, affecting a variety of crops. (Nzungize *et al.*, 2011). Symptoms of root rot are a serious threat because the damage occurs below the ground. Root rot can be caused by different groups of pathogens as fungi (Cui *et al.*, 2015).

Pythium species and Phytophthora species belong to the family Pythiaceae, both attack several crops. Symptoms include wilting, yellowing of the foliage and cause stem or root rots in numerous legumes crop, including alfalfa (Zhang and Franken, 2014). Also, alfalfa is frequently affected by *Rhizoctonia* sp. which causes root rot disease (Zhang *et al.*, 2021). *Fusarium* spp. can attack a wide range of crops. Among the symptoms, brown lesions, then become dark black lesions on below-ground roots and stems. that results in stunting and death (Subrahmanyam *et al.*, 1992).

In arid and semi-arid areas, salts frequently build up as a result of inadequate soil drainage and poor-quality irrigation water. Saline soils affect several physiological processes in plants, which reduce growth and production. While salinity stress can cause significant changes in plant physiology and morphology, little is known about how such changes may affect plant susceptibility to pathogenic organisms. The threat of increasing salinity in many irrigated areas around the world, as well as field observations, prompted some research into possible interaction between salinity stress and disease development. (Besri, 1993). Water salinity increases the population of Fusarium oxysporum in the soil, the spores of fungus in plant vessels, the formation of chlamydospores, and, finally, the development of disease even in very hot conditions. Furthermore, high salinity can break the resistance of Fusarium resistant cultivars. (Besri, 1981). Disease risk assessments have been developed and used for some soil-borne plant diseases such as F. oxysporum f. sp. lycopersici on tomato (Stirling et al., 2004), A. cochlioides on sugar beet (Almquist et al., 2016) and root rot disease of cereals (Poole et al., 2015). The aims of this study were to study the relationship between disease severity and water salinity with pathogen, salt concentration, shoot and root length.

Materials and Methods

1-Pathogenic fungi caused Alfalfa root rots

Twelve pathogenic fungal, viz, five isolates of *Fusarium* spp., two isolates of *Rhizoctonia solani*, one isolate of *Ceratobasidium* sp., one isolate of *Alternaria* sp., two isolates of *Curvularia* spp. and one isolate of *Exserohilum* sp. were obtained from the Department of Plant Pathology, Faculty of Agriculture, New Valley University, Egypt.

2-Pathogenicity tests

Pathogenicity tests of all fungi were carried out on alfalfa plants (Wadil cultivar) under greenhouse conditions in Plant Pathology Dep., Fac. of Agric., New Valley Univ. According to (Cao *et al.*, 2020) pots (25 cm in diameter) were sterilized by immersing them in 5% formalin solution for 20 minutes and left to dry before use. After that, the pots were filled with autoclaved sandy loam soil.

Alfalfa seeds were surface sterilized by dipping them in a 0.1 percent sodium hypochlorite solution for 2 minutes, then washed with sterilized water and seeded. The inoculum was grown in Erlenmeyer flasks (500 ml) with 100 ml of liquid Richard's medium in each. (McCauley *et al.*, 2005). All fungi were incubated for 10 days at 28°C before being centrifuged for 10 minutes at 3000 r.p.m. Propagules fungi were re-suspended in sterile water to give propagules concentration of 4 x 10^5 C.F.U. /ml, after one month from alfalfa seeded, fifty ml of each mycelial suspension were added to each pot. Pots with uninfected plants were used as control. Three replicates were performed for each tested isolate. After 2–months, plants were carefully uprooted and disease severity was recorded as the disease severity index (DSI) of root rots from sowing data using the arbitrary scale of 0 to 4 used by (Alejandro Rojas *et al.*, 2017).

3-Effect of saline on mycelial linear growth of pathogenic fungal in vitro

The effect of salts (NaCl or CaCO3) on the mycelial linear growth was investigated by growing the isolates in a PDA medium at three concentrations of NaCl or CaCO3 (6.25 ds/m2, 12.50 ds/m2 and 18.75 ds/m2); Control medium without added with salts. In order to test mycelial growth inhibition, a plug (5 mm) from an actively growing culture (7-10 days) was placed in the center of a PDA agar plate of 90 mm Petri dishes. Cultures were incubated at 27°C for 7-14 days, with three replications for each treatment. Colony diameters were measured at 10-15 days, as a reduction of mycelial growth (Boumaaza *et al.*, 2015). The reduction of mycelial growth percentage that occurred in each pathogenic fungus was determined at the end of the experiment using formula according to Thangavelu *et al.* (2004).

4-Effect of water salinity concentrations on Alfalfa root rots severity under greenhouse conditions

This experiment was conducted under greenhouse conditions in Plant Pathology Dep., Fac. of Agric., New Valley Univ. Inoculation methods and preparation of all fungi utilized as mentioned above. Salinity treatments tested with two salts (NaCl and CaCO3), with 3 concentrations for each salt and three replicates for each concentration. Irrigation of seedlings with 100 mL of NaCl or CaCO3 at concentrations 6.25 ds/m2, 12.50 ds/m2 and 18.75 ds/m2 (Rauf *et al.*, 2014). The control plants received SDW only. After 2–months, plants were carefully uprooted, and DS% was recorded as mentioned above.

5-The relationship between DS% with fungi, salt, root and shoot length

Depending on the experiment in greenhouse to assess the DS% of alfalfa root rots diseases under salinity conditions we studied the relationship between DS% and salinity conditions while:

DS% = dependent variable (Y)

Fungi, salt, root and shoot length = independent variable (X)

Statistical Analysis

The results of the experiments were analyzed using the software CoStat version 6.303–CoHort Software. Duncan's multiple range test was used to compare the means of all treatments at a 5% level of probability. (Gary, 2010). The STATGRAPHICS–Version 19–Stat Point, Inc. was used to prediction models. A linear model and the regression model test were used to analyze the obtained data in order to determine statistically significant differences (p = 0.05). Pearson's coefficient of correlation (r) was used to test the correlations between disease severity with fungi, salt, root and shoot length.

Results and Discussion

1-Pathogenicity tests

Data in Table (1) show that all the tested fungal isolates are significant to infect alfalfa plants (Wadi1 cv.). The tested isolates caused root rots diseases and they differed in their virulence. *Fusarium verticillioides* (F19), F. *solani* (F 39) and F. *brachygibbosum* (F23) were given the highest DS% (43.33, 45.83 and 40.00%, respectively) also recorded the highest loss of root length (58.78, 58.83 and 55.27%, respectively) and shoot length (62.77, 62.66 and 55.39%, respectively). While the lowest DS% was (24.17%) for *Exserohilum rostratum* (F10) also recorded the lowest loss of root and shoot length (21.36 and 21.38%, respectively). These results indicate that the major root rot causing agents are *Fusarium* spp. as mentioned by (Peterson *et al.*, 2018)

Fungi	Code	DS (%)	loss of root length %	loss of shoot length %
Exserohilum rostratum	F10	24.17±0.83 h*	21.36±8.861	21.38±0.66 ¹
A. alternata	F20	$27.50{\pm}1.44$ ^{gh}	$25.07{\pm}0.39^{k}$	$25.22{\pm}0.30^{k}$
C. spicifera	F25	$28.33{\pm}0.83$ gh	$28.87{\pm}0.77^{j}$	28.89 ± 0.72^{j}
C. spicifera	F38	$30.83{\pm}2.20~^{\mathrm{fg}}$	$32.77 {\pm} 0.60^{i}$	$32.95{\pm}0.67^{i}$
Ceratobasidium sp.	F7	39.17±0.83 bcd	$51.37{\pm}1.03^{d}$	51.21±1.02 ^d
Rhizoctonia solani	F26	$33.33{\pm}0.83^{defg}$	$39.98{\pm}0.35^{g}$	$39.83{\pm}0.37^{g}$
R. solani	F27	32.50±2.89 efg	$36.38{\pm}0.41^{h}$	$36.28{\pm}0.40^{h}$
F. verticillioides	F1	38.33 ± 2.20^{bcde}	47.77±0.20 ^e	47.6±10.27°
F. solani	F19	$43.33{\pm}3.63^{ab}$	58.78±1.46 ^b	58.83±1.42 ^b
F. brachygibbosum	F23	40.00 ± 1.44^{abc}	55.27±0.13°	55.39±0.05°
F. proliferatum	F30	35.83 ± 1.67^{cdef}	$43.94{\pm}2.18^{\rm f}$	43.70 ± 2.12^{f}
F. solani	F39	$45.83{\pm}3.00^{a}$	$62.77{\pm}0.40^{a}$	$62.66{\pm}0.44^{a}$
Noninfected plant (Control)		0.00 ⁱ	0.00 ^m	0.00 ^m
Mean		32.24	38.79	38.77

Table 1. Pathogenicity tests for pathogenic fungi causing alfalfa root rots diseases.

* Means followed by the same letter(s) are not significantly different (p < 0.05) according to Duncan's multiple range test.

2- Effect of salinity on mycelial growth in vitro

Table (2) illustrate that the highest significant effect of salts was detected at the Electrical Conductivity (EC value) 18.75 ds/m² which ranged (8.80 and 9.44% with NaCl and CaCO₃ reduction growth, respectively), followed by EC value 12.5 ds/m² ranged (1.13 and 0.99% reduction growth, respectively) and EC value 6.25

ds/m² (0.21 and 0.10% reduction growth, respectively). On the other hand, *Exserohilum* sp. (F10) and *A. alternata* (F20) were recorded the most affected by salinity (7.38 and 6.01% reduction, respectively). While *F. solani* (F39) was the latest affected by salinity (0.51% reduction). The other isolates were moderate significant to salinity. These results are in line with several workers (Rauf *et al.*, 2014)

					Reduction of	f mycelial gr	owth (%)		
F .					Salini	ty EC _w (ds/1	n ²)		
Fungi	code			NaCl			CaCo ₃		N
		0.00	(6.25)	(12.5)	(18.75)	(6.25)	(12.5)	(18.75)	Mean
Ex. rostratum	F10	0.00	0.96±0.49*	2.59±1.62	19.70±3.48	$0.74{\pm}0.74$	3.19±1.52	24.44±1.28	7.38±3.85ª
A. alternata	F20	0.00	0.89±6.46	2.37±0.70	18.30±3.62	0.52±0.52	1.70±0.32	18.30±2.01	6.01±3.18 ^b
C. spicifera	F25	0.00	0.67±0.67	2.22±1.28	13.19±1.55	0.00	$1.48{\pm}0.08$	14.07±2.14	4.52±2.37°
C. spicifera	F38	0.00	0.00	1.56±0.13	9.11±1.02	0.00	$1.48{\pm}0.74$	10.44±2.84	3.23±1.72 ^d
Ceratobasidium sp.	F7	0.00	0.00	0.52±0.32	4.81±0.98	0.00	0.22±0.22	5.33±1.58	$1.56{\pm}0.91^{\rm fg}$
R.solani	F26	0.00	0.00	1.26±0.20	8.67±3.02	0.00	1.04±0.52	8.59±1.28	2.79±1.52 ^{de}
R. solani	F27	0.00	0.00	1.33±0.67	9.04±1.84	0.00	1.11±0.59	8.89±1.28	2.91±1.58de
F. verticillioides	F1	0.00	0.00	0.59±0.59	5.56±1.05	0.00	0.67±0.67	5.70±0.85	1.79±1.00 ^{ef}
F. solani	F19	0.00	0.00	0.00	4.15±0.45	0.00	0.00	4.30±1.16	$1.21{\pm}0.78^{fg}$
F. brachygibbosum	F23	0.00	0.00	0.00	4.22±1.10	0.00	0.00	5.19±0.74	$1.34{\pm}0.87^{fg}$
F. proliferatum	F30	0.00	0.00	1.11±0.34	7.41±1.96	0.00	0.96±0.19	5.93±0.74	$2.20{\pm}1.18^{def}$
F. solani	F39	0.00	0.00	0.00	1.41±0.45	0.00	0.00	2.15±0.07	$0.51{\pm}0.34^{g}$
Mean		0.00 ^d	0.21±0.11 ^{cd}	1.13±0.27 ^b	8.80±1.64 ^a	$0.10{\pm}0.07^{d}$	0.99±0.27 ^{bc}	9.44±1.88 ^a	2.95±0.55

 Table 2. Effect of saline on mycelial growth of fungal pathogen in vitro causing alfalfa root rots diseases.

* Means followed by the same letter(s) are not significantly different (p < 0.05) according to Duncan's multiple range test.

3- Prediction disease severity % Models

Relationship between disease severity with fungi, salt, root and shoot length: Relationship between disease severity with fungi (Model 1)

Table (3) illustrate the linear model had the lowest percentage of correctly predicting the disease severity % expected with fungi. (r2 = 57.69%) and (r2 adjusted for d.f.= 57.49\%). In the linear model (Fig. 1). The fitted linear model's equation is:

Disease severity % Average = 22.6179 + 2.64394*Fungi

The variables' relationship is moderately strong, according to the correlation coefficient of 0.273247. Figure 2, the square root-X model had the highest accuracy in predicting the DS% expected with fungi (r2 = 76.02%) and (r2 adjusted for d.f.= 75.90\%). The square root-X model's fitted equation is:

Disease severity % Average =
$$12.0987 + 11.7261$$
*sqrt (Fungi). (1)

The variables have a moderately strong relationship, as indicated by the correlation coefficient of 0.87188.

Tab	ole 3. Re	elationship	between disease se	verity with fu	ignu						
No.	Model	Equation	Output	Correlation Coefficient	R ² %	R ² (adjusted for d.f.) %	Standard Error of Est	Mean absolute error	Durbin- Watson statistic	Lag 1 residual autocorrelation	Model No.
	Linear	$Y = a + b^*X$	DS% Average = 22.6179 + 2.64394*Fungi	0.759544	57.6907	57.4853	8.51294	6.21154	0.632082 (P=0.0000)	0.666492	ı
	Square root X	$Y = a + b^* sqrt(X)$	DS% Average = 12.0987 + 11.7261*sqrt (Fungi)	0.87188	76.0174	75.901	6.40928	5.19719	1.07714 (P=0.0000)	0.451433	
Tab	ole 4. Re	elationship) between disease se	verity with sa	alt						
No.	Model	Equation	1 Output	Correlation Coefficient	R ² %	R ² (adjusted for d.f.) %	Standard Error of Est	Mean absolute error	Durbin- Watson statistic	Lag 1 residual autocorrelation	Model No.
	Linear	Y = a + b * X	DS% Average = 22.0076 + 11.8979*Salt	0.570867 3	32.5889	32.2617	10.7455	8.57525	0.504342 (P=0.0000)	0.734772	1
7	Double square root	$Y = (a + b^* sqrt(X))$	DS% Average = (1.59116 + 3.90117*sqrt (Salt))^2	0.82214	67.5912	67.4339	1.02668	0.87263	0.5346 (P=0.0000)	0.726351	5
Tab	ole 5. Re	elationship	between disease se	verity with sł	100t leng	ţth					
No.	Model	Equation	Output	Correlation Coefficient	R ² %	R ² (adjusted for d.f.) %	Standard Error of Est	Mean absolute error	Durbin- Watson statistic	Lag 1 residual autocorrelation	Model No.
	Linear	$Y = a + b^*X$	DS% Average = 70.341 - 2.13589*Shoot average	-0.90052	81.0935	81.0017	5.69072	3.74049	0.425483 (P=0.0000)	0.781135	ı
4	Squared- Y	Y = sqrt (a + b*X)	DS% Average = sqrt (-3931.95 + 76219.8/Shoot	-0.97485	95.0336	95.0095	166.393	110.739	1.41919 (P=0.0000)	0.279997	c
			average)								

severity with fungi



severity with fungi (model 1)

Relationship between disease severity with salt (Model 2)

Table (4) shows that the linear model had the lowest percentage of correctly predicting the disease severity % expected with salt (r2 = 23.59%) and (r2 adjusted for d.f.= 32.26\%). In the linear model (Fig.3). The fitted linear model's equation is:

Disease severity % Average = 22.0076 + 11.8979*Salt

The variables' relationship is moderately strong, according to the correlation coefficient of 0.570867. On the other hand, the Double square root model had the highest accuracy in predicting the DS% expected with salt (r2 = 67.59%) and (r2 adjusted for d.f.= 67.43\%) Figure 4 shown that the Double square root model's fitted equation is:

Disease severity % Average = (1.59116 + 3.90117*sqrt (Salt)) ^2. (2)

The variables have a moderately strong relationship, as indicated by the correlation coefficient which equals 0.822139.







Tat	ole 6. Re	lationship	between disease se	verity with r	oot lengtl	h					
No.	Model	Equation	Output	Correlation Coefficient	R ² %	R ² (adjusted for d.f.) %	Standard Error of Est	Mean absolute error	Durbin- Watson statistic	Lag 1 residual autocorrelation	Model No.
v	Linear	$\begin{array}{l} Y=a+\\ b^{*}X\end{array}$	DS% Average = 70.1888 - 3.6381*Root average	-0.90111	81.1992	81.108	5.67478	3.71555	0.432562 (P=0.0000)	0.7781	
с С	Squared- Y	$\begin{array}{l} Y = sqrt \left(a + \\ b^*ln(X) \right) \end{array}$	DS% Average = sqrt (3606.7 - 224.458*Root average)	-0.9745	94.9657	94.9412	167.527	111.475	1.44207 (P=0.0000)	0.267615	4
Tat	le 7. Re	lationship	between disease se	verity with f	ungi, salt	, shoot and	root length				
						\mathbb{R}^2	Standard	Mean	Durbin-	Lao 1 residual	Model
No.	Model	Equatio	n Outj	put	R ² %	(adjusted for d.f.) %	Error of Est	absolute error	Watson statistic	autocorrelation	No.
_ د	Multiple	$Y = a + p * X_1 + b * b * X_3 + b * $	Z DS% Average X2+ 0.206622*Fungi - 17.5978*Roc X4 8.31705*Shu	c = 60.9148 - + 5.86298*Salt - ot average + oot average	88.5443	88.3186	4.46228	3.24596	0.712138 (P=0.0000)	0.641502	S
	Multiple	$Y = a + a + b^* X_1 + b^* y_3$	- DS% Average X ₂ + 0.207509*Fungi - 3.43535*Ro	c = 61.4189 - + 5.95686*Salt - ot average	88.4624	. 88.2928	4.46722	3.28288	0.659866 (P=0.0000)	0.666724	ı

Relationship between disease severity with shoot length (Model 3)

Data in Table (5) shows that the linear model had the lowest percentage of correctly predicting the disease severity % expected with shoot length (r2 = 81.09%) and (r2 adjusted for d.f.= 81.00%). in the linear model (Fig. 5). The fitted linear model's equation is:

Disease severity % Average = 70.341 - 2.13589*Shoot average

The variables' relationship is a relatively strong, according to the correlation coefficient of -0.900519. On the other hand, the the Squared-Y model had the highest accuracy in predicting the DS% expected with shoot length (r2 = 67.59%) and (r2 adjusted for d.f.= 67.43%). Figure 6 shown that the Squared-Y model's fitted equation is:

Disease severity % Average = sqrt (3618.08 - 131.91*shoot Average). (3)

The variables have a relatively strong relationship, as indicated by the correlation coefficient which equals -0.974852.



severity with shoot length

Figure 6. Relationship between disease severity with shoot length (model 3)

Relationship between disease severity with root length (Model 4)

Results in Table (6) shows that the linear model had the lowest percentage of correctly predicting the disease severity % expected with shoot length (r2 = 81.20%) and (r2 adjusted for d.f.= 81.11%). In the linear model (Fig. 7). The fitted linear model's equation is:

Disease severity % Average = 70.1888 - 3.6381*Root average

The variables' relationship is a relatively strong, according to the correlation coefficient of -0.901106. On the other hand, the the Squared-Y model had the highest accuracy in predicting the DS% expected with root length (r2 = 67.59%) and (r2 adjusted for d.f.= 67.43%). Figure 8 shown that the Squared-Y model's fitted equation is:

Disease severity % Average = sqrt (3606.7 - 224.458*Root average). (4)

The variables have a relatively strong relationship, as indicated by the correlation coefficient which equals -0.974503.



severity with root length



Relationship between disease severity with fungi, salt, shoot and root length (Models 5)

Table (7) shows that the multiple regression model had the percentage of correctly predicting the disease severity % expected with fungi, salt, shoot and root length (r2 88.54%) and (r2 adjusted for d.f.= 88.32%). In the multiple regression model (Fig. 9). The fitted multiple regression model's equation is:

Disease severity % Average = 60.9148 - 0.206622*Fungi + 5.86298*Salt - 17.5978*Root average + 8.31705*Shoot average

According to the R-Squared statistic, the fitted model accounts for 88.5443% of the variation in the DS% Average. The adjusted R-squared statistic is 88.3186%, so it is more suitable for comparing models with various numbers of independent variables.

When considering if the model can be improved keep in mind that the shoot average has the highest P-value on the independent variables at 0.2297. Because the P-value is greater than or equal to 0.05, the term is not statistically significant at the 95.0% confidence level or higher. As a result, you should think about removing shoot average from the model.

According to the result of the obtained out puts from previous model and multiple regression model had the percentage of correctly predicting the disease severity % expected with fungi, salt and root length (r2 88.4624%) and (r2 adjusted for d.f.= 88.2928%). In multiple regression (Fig. 10). The fitted multiple regression model's equation is:

Disease severity % Average = 61.4189 - 0.207509*Fungi + 5.95686*Salt - 3.43535*Root average

According to the R-Squared statistic, the fitted model accounts for 88.4624% of the variation in the DS% Average. The adjusted R-squared statistic is 88.2928%, so it is more suitable for comparing models with various numbers of independent variables.

When considering if the model can be improved keep in mind that the shoot average has the highest P-value on the independent variables at 0.2073. Because the P-value is greater than or equal to 0.05, the term is not statistically significant at the 95.0% confidence level or higher. As a result, you should think about removing fungi from the model. These results above are consistent with those reported by other researchers. (Stirling *et al.*, 2004; Poole *et al.*, 2015; Almquist *et al.*, 2016; Arafat *et al.*, 2021).



Figure 9: Relationship between disease severity with fungi, salt, shoot and root length (model 5)



Figure 10: Relationship between disease severity with fungi, salt and root length

Conclusion

Based on the study's previous results, it is concluded that using the prediction model to Prediction disease severity for alfalfa root rot. Where it helps to predict of disease severity expected if alfalfa is cultivated in an area based on soil microbial analysis and analysis of water salinity.

References

- Alejandro Rojas, J., Jacobs, J.L., Napieralski, S., Karaj, B., Bradley, C.A., Chase, T., Esker, P.D., Giesler, L.J., Jardine, D.J., and Malvick, D.K. (2017). Oomycete species associated with soybean seedlings in North America—Part I: Identification and pathogenicity characterization. Phytopathology, 107(3): 280-292.
- Almquist, C., Persson, L., Olsson, Å., Sundström, J., and Jonsson, A. (2016). Disease risk assessment of sugar beet root rot using quantitative real-time PCR analysis of *Aphanomyces cochlioides* in naturally infested soil samples. European Journal of Plant Pathology, 135:731-742.

- Arafat, K., Hassan, M., Hussein, E. (2021). Detection, disease severity and chlorophyll prediction of date palm leaf spot fungal diseases. New Valley Journal of Agricultural Science, 1(2): 98-110.
- Besri, M. (1981). Influence of irrigation water and soil salinity on the population of *Fusarium oxysporum* f. sp. *lycopersici. Phytopathologia mediterranea*.
- Besri, M. (1993). Effects of salinity on plant diseases development. In H. Lieth and A. Al Masoom eds) Towards the rational use of high salinity tolerant plants, 2: 67-74. Kluwer Academic Publishers.
- Bingham, E., Hurley, L., Kaatz, D., and Saunders, J. (1975). Breeding Alfalfa Which Regenerates from Callus Tissue in Culture 1. Crop Science, 15(5): 719-721.
- Boumaaza, B., Benkhelifa, M., and Belkhoudja, M. (2015). Effects of two salts compounds on mycelial growth, sporulation, and spore germination of six isolates of *Botrytis cinerea* in the western north of Algeria. International Journal of Microbiology, 2015 :572626.
- Cao, S., Liang, Q., Nzabanita, C., and Li, Y. (2020). Paraphoma root rot of alfalfa (*Medicago sativa*) in Inner Mongolia, China. Plant pathology, 69(2): 231-239.
- CAPMAS (2023). Central Agency for Public Mobilization and Statistics, database. (<u>https://www.capmas.gov.eg</u>).
- Conrad, H., and Klopfenstein, T. (1988). Role in livestock feeding—Greenchop, silage, hay, and dehy. Alfalfa and alfalfa improvement, 29: 539-551.
- Cui, B.-K., Dai, Y.-C., He, S.-H., Zhou, L.-W., and Yuan, H.-S. (2015). A novel *Phellinidium* sp. causes laminated root rot on Qilian juniper (*Sabina przewalskii*) in northwest China. Plant disease, 99(1): 39-43.
- Gary, W. O. (2010). A first course in design and analysis of experiments. University of Minnesota, Minnesota, United States of America.
- Harrison, M. J., Dewbre, G. R., and Liu, J. (2002). A phosphate transporter from Medicago truncatula involved in the acquisition of phosphate released by arbuscular mycorrhizal fungi. The Plant Cell, 14(10): 2413-2429.
- McCauley, A., Jones, C., and Jacobsen, J. (2005). Basic soil properties. Soil and water management module, 1(1): 1-12.
- Nzungize, J., Gepts, P., Buruchara, R., Male, A., Ragama, P., Busogoro, J., and Baudoin, J.-P. (2011). Introgression of Pythium root rot resistance gene into Rwandan susceptible common bean cultivars. African Journal of Plant Science, 5(3): 193-200.
- Peterson, J., Samac, D., and Grau, C. (2018). First Report of Fusarium Wilt of Alfalfa Caused by *Fusarium oxysporum* f. sp. *medicaginis* in Wisconsin. Plant disease, 102(2): 447-447.
- Poole, G. J., Harries, M., Hüberli, D., Miyan, S., MacLeod, W., Lawes, R., and McKay, A. (2015). Poole, G. J., Harries, M., Hüberli, D., Miyan, S., MacLeod, W., Lawes, R., and McKay, A. (2015). Predicting cereal root disease in Western Australia using soil DNA and environmental parameters. Phytopathology, 105(8): 1069-1079.
- Rauf, A., Zaki, M. J., and Khan, D. (2014). Effects of NaCl salinity on growth of some cotton varieties and the root rot pathogens. Int. J. Biol. Biotech, 11(4): 661-670.

- Stirling, G., Griffin, D., Ophel-Keller, K., McKay, A., Hartley, D., Currar, J., Hardie, B. (2004). Combining an initial risk assessment process with DNA assays to improve prediction of soilborne diseases caused by root-knot nematode (*Meloidogyne* spp.) and *Fusarium oxysporum* f. sp. *lycopersici* in the Queensland tomato industry. Australasian Plant Pathology, 33(2): 285-293.
- Subrahmanyam, P., Wongkaew, S., Reddy, D., Demski, J. W., McDonald, D., Sharma, S., and Smith, D. (1992). Field diagnosis of groundnut diseases.Monograph. International Crops Research Institute for the Semi-Arid Tropics. (<u>http://oar.icrisat.org/id/eprint/1227</u>)
- Thangavelu, R., Sundararaju, P., and Sathiamoorthy, S. (2004). Management of anthracnose disease of banana caused by *Colletotrichum musae* using plant extracts. The Journal of Horticultural Science and Biotechnology, 79(4): 664-668.
- Van Keuren, R., Matches, A., Hanson, A., Barnes, D., and Hill JR, R. (1988). Alfalfa and alfalfa improvement. Series Agronomy. Pasture production and utilization, 29: 515-538.
- Zhang, C., Yu, S., Tian, H., Wang, Z., Yu, B., Ma, L., Nan, Z., and Fang, X. (2021). Varieties with a high level of resistance provide an opportunity to manage root rot caused by *Rhizoctonia solani* in alfalfa. European Journal of Plant Pathology, 1-7.
- Zhang, H., and Franken, P. (2014). Comparison of systemic and local interactions between the arbuscular mycorrhizal fungus *Funneliformis mosseae* and the root pathogen *Aphanomyces euteiches* in *Medicago truncatula*. Mycorrhiza, 24(6): 419-430.

التنبؤ بشدة الإصابة بأمراض أعفان جذور البرسيم الحجازي تحت الظروف الملحية

خالد حسين عرفات¹*، محمد حسن عبدالرحيم حسن²، عمر حسين حسن محمد¹

¹قسم امراض النبات، كلية الزراعة، جامعة الوادي الجديد، مصر. ²قسم امراض النبات، كلية الزراعة، جامعة اسيوط، مصر.

الملخص

تصاب جذور البرسيم الحجازي بالعديد من مسببات الأمراض الفطرية تحت الظروف الملحية. تم در اسة تأثير ثلاث تركيز ات من ملحين مختلفين هما كلوريد الصوديوم وكربونات الكالسيوم على نمو الفطريات المسببة لاعفان جذور البرسيم الحجازي في المعمل وكذلك تأثير هذه الاملاح على شدة الإصابة على النباتات في الصوبة لايجاد العلاقة بين شدة الإصابة بالفطريات الممرضة وملوحة مياه الري. معمليا: وجد ان أعلى تأثير على نمو الميسليوم الفطري تم الحصول عليه عند تركيز 18.75 ديسيسيمنز /م2 حيث تراوح من (8.80 و 9,44 % مع كلوريد المسوديوم وكربونات الكالسيوم على التوالي. علاوة على ذلك فطر Exserohilum sp. كان اعلى الفطريات تأثرا ملوحة المياه. مع المستويات المختلفة. وجد ان شدة الإصابة بأعفان الجذور تزداد بزيادة التركيزات المختلفة من الملوحة. وأدى التركيز الأعلى للملوحة (18.75 ديسيسيسيمنز /م2) لكلا الملحين الى زيادة شدة الإصابة حيث تراوحت من (42.76، 43.53% بالنسبة لكلوريد الصوديوم و44.04، 44.42 % بالنسبة لكربونات الكالسيوم). ومن ناحية اخرى مع التركيزات المختلفة لكلا الملحين حصل الفطر Fusarium sp. على اعلى متوسط لشدة الإصبابة وبذلك بالاعتماد على النتائج التي تم الحصبول عليها أمكن تصميم نموذج تنبؤ لتوضيح العلاقة بين الملوحة وشدة الإصبابة وطول الجذر والسباق، حيث وجد أن معامل ارتباط العلاقة بين النسبة المئوية لشدة الإصبابة بالفطريات ونوع وتركيز الملح وطول الجذر والسباق في نموذج الانحدار المتعدد هي 88.83% . تهدف هذه الدر اسـة الي ايجاد العلاقة بين قيم شـدة الإصـابة وتركيز ات الملوحة للتنبؤ المبكر بشدة الإصابة المرضبة باعقان الجذور