

## Assessment of Genetic Diversity Using SCoT Markers and Some Morphological Traits in Ten Lines of Barley (*Hordeum vulgare* L.)

\* Habiba, Rehab M.<sup>1</sup>; Jamila Bashasha<sup>2</sup>; Soad H. Haffez<sup>3</sup> and A.A.A. Abo Leilah<sup>3</sup>

<sup>1</sup>Department of Genetics, Faculty of Agriculture, Mansoura Univ., Mansoura 35516, Egypt

<sup>2</sup>Benghazi Univ., Faculty of Agriculture, Horticulture Department, Libya

<sup>3</sup>Department of Agronomy, Faculty of Agriculture, Mansoura Univ., Mansoura 35516, Egypt

\* Corresponding author; E-mail address: [rehab74@mans.edu.eg](mailto:rehab74@mans.edu.eg)

Accepted for publication on: 31/12/2021



### Abstract

To assessment molecular and phenotypic diversity for ten barley lines belong to *Hordeum vulgare* L., ten SCoT primers were used and 12 morphological traits were estimated in two seasons (2018/2019 and 2019/2020). The SCoT primers succeeded in generating reproducible and reliable amplicons. SCoT technique showed that 66.67 % to 100% of polymorphism. The resolving power (Rp) value varied from 4 to 11.40. In addition, the 10 lines were characterized by 41 unique markers (22 positive and 19 negative). B<sub>6</sub> had the highest numbers of positive markers (six). According to phenotypic evaluation, the mean squares for genotypes were highly significant for all studied traits from combined data over two seasons. The heritability values in broad sense ( $h^2_b$  %) ranged from 40.63 (100-grain weight) to 99.22 (Days to heading). The P<sub>7</sub> gave desired value in four traits (NT/P, NS/P, NG/S and GY/P g) and the other lines showed desired value in one or two trait, thus all traits which detected in the ten lines might be associated with all unique markers distinguished in this study. The inbred line P<sub>6</sub> showed the highest number of unique markers (6 positive), one or some of which may be linked with grain filling period (GFP day) trait that showed in line desirable value. Consequently, these markers may be used as selectable markers for genetic improvement of these traits in barley.

**Keywords:** SCoT technique, barley, Genetic diversity, Molecular distance, Cluster analysis.

### Introduction

Barley (*Hordeum vulgare*) was one of the primary cultivated grains, as early as 10,000 years ago. It is a major cereal grain grown in temperate climates globally, self-pollinated and diploid ( $2n = 14$ ). In the ancient, barley flour was the main used for breads. Malt is used to produce purified alcohol, malt syrup and malted milk. Co-products from malting and brewing are also used in fodder manufacture.

Assessment of genetic diversity and phenotyping of germplasm provide information about traits variability in any crop species and offers a

basis for planning future strategies for crop development. Thus, used of molecular markers via marker-assisted selection may be improve the main traits in barley such as yield components traits. The relationship between molecular markers and morphological trait evaluation is one of the most important factors in the plant breeding and molecular genetics. It delivers important landmarks for exposition of genetic variability and discovery of genomic regions that are responsible for the morphological trait, which plays an important role in the development of barley using marker-assisted selection Dora *et al.* (2017)

and Adawy *et al.* (2008). In barley, The genetic diversity has been investigated using diverse molecular techniques as SCoT, ISSR and RAPD markers such as Aboulila and Mansour (2017), Amer *et al.* (2017), Dora *et al.* (2017) and Fernandez *et al.*, (2002). SCoT is one of molecular marker system described by Collard and Mackill (2009). It built on the small preserved regions of genes are flanked by the ATG start codon of translation. SCoT uses single primer designed to anneal the around regions of the ATG on the two strands of DNA. SCoT technique is dominant marker like RAPDs (Xiong *et al.*, 2011) and might be used for genetic analysis, QTL mapping and bulk segregation analysis.

The present study aimed to study the genetic diversity in ten barley lines at molecular and phenotypic levels using SCOT molecular markers. Thus, it would be determine the molecular markers that can be linked with distinguished traits in each studied line.

#### **Materials and Methods**

This study was carried out at the molecular genetic laboratory Genetics Department and Experimental Farm at Agronomy Department, Faculty of Agriculture, Mansoura University, Mansoura, Egypt. The plant material used in this study comprised of ten Libyan local lines of barley belong to species *Hordeum vulgare* were supplied from the Faculty of Agriculture, Benghazi Univ.

#### **Molecular evaluation:**

DNA was isolated from fresh leaves by DNeasy plant mini kit (QIAGEN). Genomic DNA was used as a template for Polymerase Chain Reaction using ten SCoT primers in molecular evaluation for 10 lines of barley. Amplification reactions for

SCoT technique were carried out in Techni TC-512 Thermal Cycler according to Abd El-Aziz *et al.* (2019). The PCR cycles were carried out according to Rehab *et al.* (2020). DNA profiles were photographed using Bio-1D Gel Documentation system and analyzed by Gel Analyzer 3 software. DNA-profiles were done for SCoT technique according to Adhikari *et al.* (2015). Polymorphic Information Content (PIC) and Diversity Index (DI) were calculated according to,  $PIC = 1 - p^2 - q^2$  (Gorji *et al.*, 2011). In addition, the capability of each primer to distinguish between lines was assessed according to resolving power value (Rp) calculated as described in Prevost and Wilkinson (1999). Molecular distances (MD) were calculated by Dice coefficient (Nei and Li, 1979) and cluster analysis was done using XLSTAT.7 software.

#### **Phenotypic evaluation:**

A two-year field trial was conducted at Experimental Farm, Agronomy Department. During two successive growing seasons, 2018/2019 and 2019/2020, ten barley lines were evaluated in a randomized complete block design (RCBD) with three replications. Lines were evaluated using 12 agro-morphological traits. Analysis of variance was employed in order to test the significance of the difference between the lines for the various traits across the two years. In addition, a combined analysis of variance for genotypes over the two years was made for the studied traits according to Steel and Torrie (1960). The least significant difference (LSD) test for mean comparisons was down using SAS (Ver. 9.1). The data were recorded on ten guarded randomly chosen plants per plot for all genotypes. The same procedure was followed in

the two seasons. The studied traits were: Plant height (PH cm), Number of tillers (NT/P), Number of spikes/plant (S/P), Spike length (SL cm), Number of grains/spike (NG/S), 100-grain weight (100/GWT g), Flag leaf area (LA cm<sup>2</sup>), Days to heading (DH day), Days to maturity (DM day), Grain filling period (GFP day), Grain filling rate (GFR g/day) and Grain yield per plant (GY/P g). Finally, the heritability was estimated on the basis of entry mean following  $\sigma^2_g / (\sigma^2_g + \sigma^2_{gy/ry} + \sigma^2_{e/r})$ , where  $\sigma^2_g$ ,  $\sigma^2_{gy}$  are the variance component due to genotypes, genotypes by years interaction and  $\sigma^2_e$  is variance components due to unexplainable error.  $r$ ,  $y$  are the number of replicates and years.

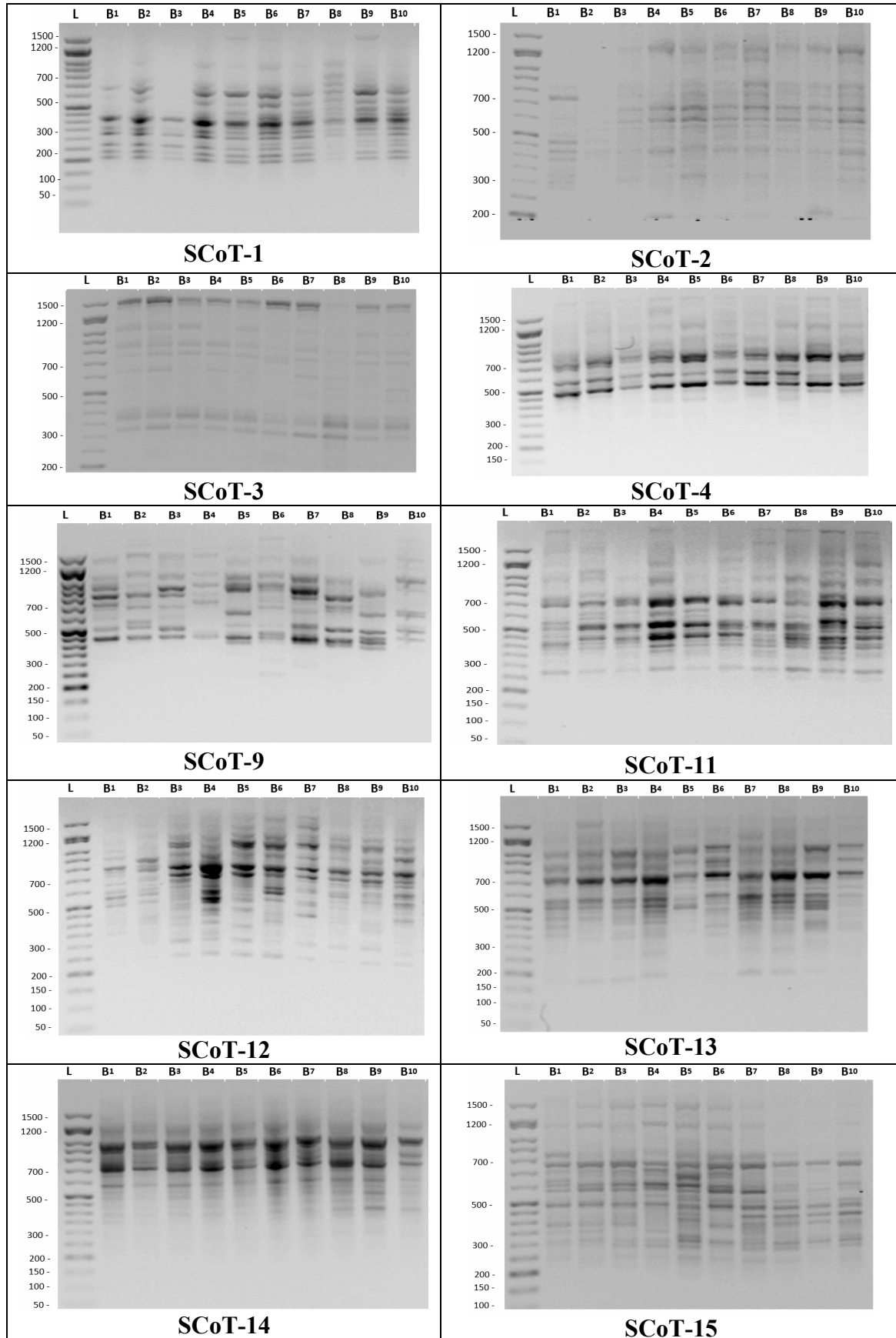
### Results and Discussion

Ten SCoT primers were used to study the genetic differences and relationships among the 10 barley genotypes as shown in Fig. 1 and Table 1. The molecular sizes of the amplified bands varied with the different SCoT primers and ranged from 182 bp to 2203 bp. All these SCoT primers generated good banding patterns that indicated the influence of SCoT marker in fingerprinting and genetic diversity.

A total of 180 major SCoT amplified fragments were produced, out of them 143 (79.44%) were polymorphic and the polymorphism percentage ranged from 66.67 % (SCoT-1) to 100% (SCoT-9). Similarly, high level of polymorphism in barley was founded by Amer *et al.* (2017) and Dora *et al.* (2017) who found that level of polymorphism ranged from 55.56% to 91.67%. In tomato genotypes, polymorphism was 100% founded by Henareh *et al.* (2016) and

80 to 100% founded by Rehab *et al.* (2020). While, low level of polymorphism was found by Shahlai *et al.* (2014) (23.25%). The total number of polymorphic fragments of DNA ranged from low scored by the two primers SCoT-1 and SCoT-3 (10), to high scored by the primer SCoT-12 (22). All these primers generated good banding patterns that illustrated the rule of SCoT marker in fingerprinting and diversity analyses. These differences in the number of amplified bands by altered SCoT primers are influenced by variable reasons such as number of annealing sites in the genome and primer structure (Kernodle *et al.*, 1993).

The polymorphism index content (PIC) analysis was carried out to conclude the efficiency of each primer SCoT to express polymorphic loci in barley. The calculated PIC values for primers SCoT ranged from 0.222 to 0.348. The primer SCoT-3, which produced the lowest mean PIC value of 0.222, was the least polymorphic. Dora *et al.* (2017) found that the PIC ranged from 0.18 to 0.33. Resolving power (RP) is used to describe the capacity of the primer/marker combination to detect the differences among various genotypes (Prevost and Wilkinson, 1999). RP values of the ten SCoT primers ranged from 4 to 11.40 characteristic the different genotypes whereas the average was 7.76 per SCoT primer. The highest RP values were detected with the primer SCoT-12 (11.40) and the lowest with the primer SCoT-3 (4). In this respect, Dora *et al.* (2017) found that the RP values for SCoT markers was ranged from 9.6 to 12.7 in barley.



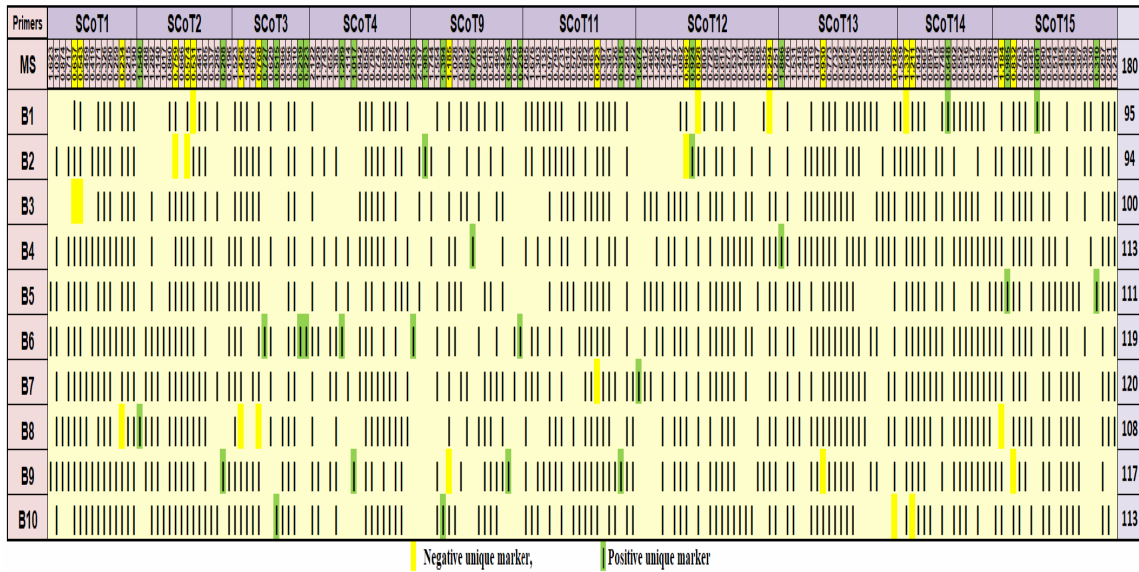
**Fig. 1:** Banding patterns of SCoT-PCR products for ten barley lines, M, 1.5 K bp ladder and lanes 2 to 11 represent the ten genotypes.

**Table 1: Molecular data estimated from banding patterns of SCoT technique.**

Primer		Amplicons						Polymorphism %	Polymorphic index content (PIC)	Resolving power Rp
Name	Sequence (5' → 3')	Molecular size range	Monomorphic	Polymorphic			Total			
				Without unique	Unique +	Unique -				
SCoT-1	CAACAATGGCTACCACCA	190:1623	5	7	-	3	15	66.67	0.231	5.00
SCoT-2	CAACAATGGCTACCACCC	196:1540	1	10	2	3	16	93.75	0.316	7.80
SCoT-3	CAACAATGGCTACCACCG	223:1523	3	4	4	2	13	76.92	0.222	4.00
SCoT-4	CAACAATGGCTACCACCT	454:2122	5	10	2	-	17	70.59	0.278	7.60
SCoT-9	CAACAATGGCTACCAGCA	239:2203	-	12	6	1	19	100.00	0.348	10.20
SCoT-11	AAGCAATGGCTACCACCA	222:2065	5	12	1	1	19	73.68	0.306	9.40
SCoT-12	ACGACATGGCTACCAACG	251:1674	2	17	2	3	24	91.67	0.324	11.40
SCoT-13	ACGACATGGCGACCATCG	182:1866	6	11	1	2	20	70.00	0.278	8.80
SCoT-14	ACGACATGGCGACCACGC	286:1539	5	8	1	2	16	73.68	0.234	5.40
SCoT-15	ACGACATGGCGACCGCGA	244:1519	5	11	3	2	21	76.19	0.257	8.00
<b>Overall</b>		<b>182:2203</b>	<b>32</b>	<b>102</b>	<b>22</b>	<b>19</b>	<b>180</b>	<b>79.32</b>	<b>0.28</b>	<b>7.76</b>

The genetic fingerprints for all ten lines of barley were performed as DNA-profile diagram (Figure 2) based on 180 amplicons obtained using 10 SCoT primers. This profile showed that the amplicons per lines were variously ranged from 94 (for B<sub>1</sub>) to 120 (for B<sub>7</sub>). In addition, the 10 lines were categorized by 41 unique bands (markers) (22 positive and 19 negative). B<sub>6</sub> had the highest numbers of positive bands (six). These markers were spread over these lines variously differentiate each lines from the other. These unique bands may be suitable as unique markers as clarified by Abd El-Aziz *et al.* (2016)

and Rehab *et al.* (2020) in tomato, Abd El-Aziz and Rehab (2016) in canola and Abd El-Hadi *et al.* (2017) in squash. These results indicated that DNA-profiling diagram also is a good tool for molecular identification for ten barley genotypes. Thus, it was assumed that SCoT primers used in this study were with high degree of confidence for the molecular identification. These results agree with Aboulila and Mansour (2017) who studied the genetic diversity between ten barley genotypes using SCoT marker, and they described that SCoT marker is an effective tool for finding new fingerprint of barley.



**Fig. 2:** DNA-profile representations of SCoT fingerprint of ten barley lines based on 180 amplicons 41 of them were marker loci.

According to Table 3, the molecular distance (MD) among all studied lines based on SCoT data ranged from 0.447 to 0.839. The highest molecular distance (MD) was among B<sub>9</sub> and B<sub>2</sub> (0.839) followed by B<sub>5</sub> and B<sub>1</sub> (0.830). While the lowest MD was

between B<sub>8</sub> and B<sub>7</sub> (0.447) followed by B<sub>10</sub> and B<sub>9</sub> (0.529). This means that B<sub>9</sub> and B<sub>2</sub> were the best genotypes that can be used in breeding programs to obtain the genetic variability and/or hybrid vigor from hybridization between them.

**Table 3. The molecular distance (MD) between all ten barley lines based on SCoT data.**

	B1	B2	B3	B4	B5	B6	B7	B8	B9
B2	0.590								
B3	0.614	0.535							
B4	0.738	0.609	0.532						
B5	0.830	0.725	0.532	0.638					
B6	0.737	0.815	0.601	0.677	0.695				
B7	0.630	0.780	0.613	0.594	0.539	0.585			
B8	0.705	0.714	0.680	0.690	0.571	0.698	0.447		
B9	0.738	0.839	0.766	0.818	0.596	0.615	0.605	0.553	
B10	0.784	0.747	0.671	0.737	0.567	0.604	0.569	0.573	0.529

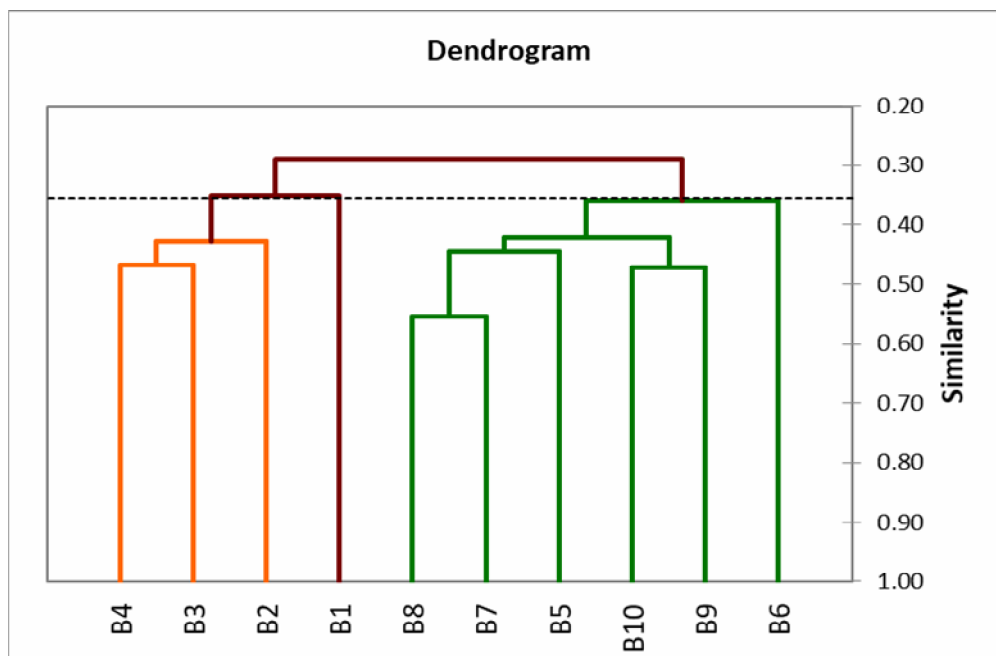
According to the UPGMA clustering algorithm from SCoT markers, the 10 lines of barley were divided into three major clusters (Fig. 3). The first major cluster was divided into two sub-clusters. The first sub-cluster consisted of lines B<sub>4</sub> and B<sub>3</sub> and the second sub-cluster consisted of lines

B<sub>2</sub>. The second main cluster consisted of one lines B<sub>1</sub>. In addition, the third main cluster was divided into two sub-clusters. The first sub-cluster contained of lines B<sub>8</sub>, B<sub>7</sub>, B<sub>5</sub>, B<sub>10</sub> and B<sub>9</sub> and the second sub-cluster contained of one line B<sub>6</sub> which was substantiated before by the presence of

highest number of specific unique bands over other tested lines.

In conclusion, this study confirms the capability of SCoT as a good marker to examine the genetic relationships between different lines

of barley and obtaining new specific markers. Documentation of new specific markers is very significant for breeders to evaluate barley genotypes for breeding programs.



**Fig.3:** Dendrogram derived by UPGMA method using Dice-similarity coefficient for binary data of SCoT technique for ten barley lines.

Legend: TL represents truncated line at a coefficient of Similarity=0.35

### Phenotypic evaluation:

The combined analyses of variance for the agronomic traits are shown in Table 4. The obtained results revealed that the magnitudes of the mean squares for genotypes were highly significant for all studied traits, indicating the presence of genetic differences among barley genotypes for the studied traits. These result in agreement with Pesaraklu *et al.* (2016). As well as, the mean squares due to genotypes by year interaction were highly significant for plant height (PH cm), number of tillers (NT/P), and number of spikes/plant (NS/P). The mean squares due to years were highly sig-

nificant for number of tillers (NT/P) and number of spikes/plant (NS/P). While, it was significant for plant height (PH cm) and Spike length (SL cm). These results suggest that these lines of barley showed different performances at different environments conditions and may have different combining ability pattern and in crosses depending on type of other parent. In addition, the heritability values in broad sense ( $h^2_b$  %) ranged from 40.63 (100-grain weight) to 99.22 (Days to heading). Pesaraklu *et al.* (2016) found that the heritability values in broad sense ( $h^2_b$  %) ranged from 47.7 (Plant height) to 87.7 % (Grains per spike)

**Table 4. Estimated mean squares and broad-sense heritability ( $h^2_b$  %) of different agronomic traits in ten barley lines from the combined data over two growing seasons**

S.O.V	d.f	PH (cm)	NT/P	NS/P	SL (cm)	NG/S	100/GWT
Rep/Years	4	60.01**	6.07	3.27	0.77	111.03	0.12
Years (Y)	1	38.40*	70.42**	62.02**	1.66*	0.00	0.01
Genotypes(G)	9	418.67**	52.83**	59.68**	7.11**	687.42**	0.97**
G/Y	9	90.94**	22.42**	21.31**	0.44	0.000	0.45
Error	36	12.39	4.03	3.58	0.38	87.89	0.31
$h^2_b$ %		76.03	53.48	60.65	89.05	79.64	40.63
S.O.V	d.f	LA	DH day	DM day	GFP	GFR	GY/P
Rep/Years	4	2.12	0.93	1.38	4.27	0.01	7.19
Years (Y)	1	0.51	0.00	20.42	24.07	0.008	0.001
Genotypes(G)	9	484.71**	752.96**	464.91**	252.51**	0.47**	1146.46**
G/Y	9	0.17	3.11	1.75	4.92	0.003	5.52
Error	36	15.16	2.80	2.88	6.03	0.01	9.13
$h^2_b$ %		94.11	99.22	98.77	95.36	95.89	98.42

Plant height (PH cm), Number of tillers (NT/P), Number of spikes/plant( S/P), Spike length (SL cm), Number of grains/spike (NG/S), 100-grain weight (100/GWT g), Flag leaf area (LA cm<sup>2</sup>), Days to heading (DH day), Days to maturity (DM day), Grain filling period (GFP day), Grain filling rate (GFR g/day) and Grain yield per plant (GY/P g).

The results from Table 5 showed the lowest and highest mean performance values for studied phenotypic traits of ten barley lines. In all traits, the favorable genotypes were

the ones with higher values of the traits except for plant height, days to heading, days to maturity, grain filling period and grain filling rate.

**Table 5. Mean performance range and desirable values for all studied traits in ten lines**

desirable value	High		Low		Traits
	Lines	Value	Lines	Value	
Low	B <sub>6</sub>	124.41	B <sub>3</sub>	99.93	PH cm
High	B <sub>7</sub>	30.00	B <sub>8</sub>	20.00	NT/P
High	B <sub>7</sub>	28.67	B <sub>8</sub>	18.67	NS/P
High	B <sub>1</sub>	11.67	B <sub>10</sub>	8.17	SL cm
High	B <sub>7</sub>	88.00	B <sub>6</sub>	52.66	NG/S
High	B <sub>5</sub>	7.44	B <sub>9</sub>	6.10	100/GWT g
High	B <sub>10</sub>	51.55	B <sub>2</sub>	22.13	LA cm <sup>2</sup>
Low	B <sub>7</sub>	95.00	B <sub>3</sub>	62.33	DH day
Low	B <sub>10</sub>	156.50	B <sub>2</sub>	131.33	DM day
Low	B <sub>3</sub>	73.50	B <sub>6</sub>	53.00	GFP day
Low	B <sub>7</sub>	1.66	B <sub>8</sub>	0.04	GFR g/day
High	B <sub>7</sub>	91.90	B <sub>8</sub>	39.45	GY/P g

Plant height (PH cm), Number of tillers (NT/P), Number of spikes/plant( NS/P), Spike length (SL cm), Number of grains/spike (NG/S), 100-grain weight (100/GWT g), Flag leaf area (LA cm<sup>2</sup>), Days to heading (DH day), Days to maturity (DM day), Grain filling period (GFP day), Grain filling rate (GFR g/day) and Grain yield per plant (GY/P g).



In the present study, the tested SCoT primers showed high efficiency in generating specific unique markers for all tested lines of barley. Based on desirable performance of ten lines for studied traits, data presented in Table 6 clearly reflected that plant height (PH cm), number of tillers (NT/P), number of spikes/plant (NS/P), spike length (SL cm), number of grains/spike (NG/S), 100-grain weight (100/GWT g), flag leaf area (LA cm<sup>2</sup>), days to heading (DH day), days to maturity (DM day), grain filling period (GFP day), grain filling rate (GFR g/day) and grain yield per plant (GY/P g) traits which showed desirable values could be associated with some unique markers detected in this study. These unique markers were 19 negative and 22 positive SCoT DNA fragments. The line P<sub>6</sub>

showed the highest number (six) of positive unique markers where one or more of them may be linked with grain filling period (GFP day) trait that indicated in this line a desirable value. Similarly, the line B<sub>1</sub> which exposed two positive unique markers at least one of them may be linked with spike length (SL cm) trait. Also, B<sub>5</sub> showed two positive unique markers one or more of them may be associated with 100-grain weight (100/GWT g) trait. These results illustrated that some of these markers may be used as markers assisting selection in the breeding program to improve barley lines. Similar conclusion was obtained by Giancarla *et al.* (2012) in barley, Abd El-Aziz *et al.* (2017) in okra and Abd El-Hadi *et al.* (2017) in squash.

**Table 6: The relationship between molecular markers and distinguished traits**

Distinguished traits		Unique markers				Inbred lines
Mean performance	Trait	Total	Type	Molecular size	Primer	
11.67	SL cm	6	- - + - +	531 294,828 648 1337 310	SCoT-2 SCoT-12 SCoT-14 SCoT-14 SCoT-15	B <sub>1</sub>
131.33	DM day	5	- - + -	574,759 1983 924 962	SCoT-2 SCoT-9 SCoT-12 SCoT-12	B <sub>2</sub>
99.93 62.33	PH cm DH day	2	-	543,637	SCoT-1	B <sub>3</sub>
---	--	2	+ +	776 1866	SCoT-9 SCoT-13	B <sub>4</sub>
7.44	100/GWT g	2	+	310, 985	SCoT-15	B <sub>5</sub>
53.00	GFP day	6	+ + +	223,239,727 1209 239, 2203	SCoT-3 SCoT-4 SCoT-9	B <sub>6</sub>
30.00 28.67 88.00 91.90	NT/P NS/P NG/S GY/P g	2	- +	423 1674	SCoT-11 SCoT-12	B <sub>7</sub>
0.04	GFR g/day	5	- + - -	234 1540 768,1426 1184	SCoT-1 SCoT-2 SCoT-3 SCoT-15	B <sub>8</sub>
---	---	7	+ - + - + - -	208 1017 354 1185 315 930 832	SCoT-2 SCoT-4 SCoT-9 SCoT-9 SCoT-11 SCoT-13 SCoT-15	B <sub>9</sub>
51.55	LA cm <sup>2</sup>	4	+ + - -	515 1394 182 1211	SCoT-3 SCoT-9 SCoT-13 SCoT-14	B <sub>10</sub>

Plant height (PH cm), Number of tillers (NT/P), Number of spikes/plant( NS/P), Spike length (SL cm), Number of grains/spike (NG/S), 100-grain weight (100/GWT g), Flag leaf area (LA cm<sup>2</sup>), Days to heading (DH day), Days to maturity (DM day), Grain filling period (GFP day), Grain filling rate (GFR g/day) and Grain yield per plant (GY/P g).

## References

- Abd El-Aziz M. H.; S. Y. Mohamed and Hadeer E. Magwaid (2019). Molecular and phytochemical assessment for some seedy strains of Alamar apricot rootstock under salinity stress. *Egyptian Journal of Basic and Applied Sciences*, 6 (1): 173–186.
- Abd El-Aziz, M. H. and Rehab M. M. Habiba (2016). Molecular assessment of genetic diversity in some canola homozygous lines. *Egypt J. Genet. Cytol.*, 45: 129-145.
- Abd El-Aziz, M. H.; K. A. Zaied; Soher E. A. El-Gendy and N. A. Abd El-Gawad (2017). Evaluation of irradiated okra based on agronomical traits and RAPD Markers. *Assiut J. Agric. Sci.*, 48 (3): 81-96.
- Abd El-Aziz, M. H.; S.M. Farid and Sara A.A. Elkomey (2016). Evaluation of molecular and phenotypic diversity in relation to heterosis in some tomato lines under different climatic conditions. *J. Agric. Chem. and Biotechn., Mansoura Univ.*, 7 (5): 141- 151.
- Abd El-Hadi, A. H.; M. H. Abd El-Aziz; Manal A. Abd Alla and Mariam G. Ashak (2017). Molecular and Phenotypic Evaluation of some Summer Squash Inbred Lines *J. Agric. Chem. and Biotechn., Mansoura Univ.* Vol. 8 (12): 281 – 287.
- Aboulila, A.A. and M. Mansour (2017). Efficiency of Triple-SCoT primer in characterization of genetic diversity and genotype-specific markers against SSR fingerprint in some Egyptian barley genotypes. *Am. J. Mol. Biol.* 7(03): 123-137.
- Adawy, S.; M. Saker; W. Hagag and H. El Itriby (2008). AFLP-based molecular analysis of Egyptian barley lines and landraces differing in their resistance and susceptibility to leaf rust and net blotch diseases. *J. of Landbauforschung*, 58: 125-134.
- Adhikari, S.; S. Saha and TK. Bandyopadhyay (2015). Efficiency of ISSR marker for characterization of *Cymbopogon* germplasm and their suitability in molecular barcoding. *Plant Syst. Evaluation.* 301:439 – 450.
- Amer, Kh. A.; R.A. Abou El Enein; A.A. El-Sayed; M.M. Noaman *et al.* (2017). Giza 137 and Giza 138, new Egyptian six-rowed barley cultivars for new land. *Egypt. J. Plant Breed.* 21 (5): 380 – 395.
- Collard, B. C. Y. and D. J. Mackill (2009). Start codon targeted (SCoT) polymorphism: a simple, novel DNA marker technique for generating gene-targeted markers in plants. *Plant Mol. Biol. Rep.* 27, 86–93.
- Dora, S.A; M. Mansour; A.A. Aboulila and E. Abdelwahab (2017). Genetic Diversity and Relationships among Some Barley Genotypes for Net Blotch Disease Resistance Using RAPD, SCoT and SSR Markers. *Egypt. J. Genet. Cytol.*, 46: 139-165,
- Fernandez, M. N.; A. M. Figueiras and C. Benito (2002). The use of ISSR and RAPD markers for detection DNA polymorphism, genotype identification and genetic diversity among barley cultivars with known origin. *Theor. Appl. Genet.*; 104: 845–851.
- Giancarla, V.; M. Emilian; Ş. Radu; C. Soriin; P. Sorina and P. Cerasela (2012). The use of RAPD and ISSR markers for genetic diversity among some barley cultivars. *Romanian Biotechnological Letters*, 17(4): 7493 -7503.
- Gorji, A. M., P. Poczai, Z. Polgar and J. Taller (2011). Efficiency of arbitrarily amplified dominant markers (SCoT, ISSR and RAPD) for diag-

- nostic fingerprinting in tetraploid potato. *Am. J. Pot. Res.*, 88: 226-237.
- Henareh, M.; A. Dursun; B. Abdollahi Mandoulakani and K. Haliloğlu (2016). Assessment of genetic diversity in tomato landraces using ISSR markers. *Genetika*, Vol. 48 (1): 25-35.
- Kernodle, S. P.; R. E. Cannon and J. G. Scandalios (1993). Concentration of primer and template qualitatively affects product in RAPD PCR. *Biotechniques*, 1: 362-364.
- Nei, M. and W.H. Li (1979). Mathematical model for studying genetic variation in terms of restriction endonucleases. *Proc Natl Acad Sci, USA.*; 76: 5269–5273
- Pesaraklu, S.; H. Soltanloo; S.S Ramez-anpour.; M. KalateArabi and Nasrollah NejadGhomi A. A. (2016). An estimation of the combining ability of barley genotypes and heterosis for some quantitative traits. *Iran Agricultural Research*, 35(1): 73-80
- Prevost, A. and M.J. Wilkinson (1999). A new system of comparing PCR primers applied to ISSR fingerprinting of potato cultivars. *Theo. App. Genet.*; 98:107–112.
- Rehab M. Habiba; A.Y.A.M. Elsayed and Hasnaa A. Saleh (2020). Enhancement of Chilling Tolerance and Efficiency Using Start Codon Targeted (SCoT) Technique for Molecular Assessment in some Tomato Genotypes. *J. of Agric. Chem. and Biotechn.*, Mansoura Univ. Vol. 11(10): 285- 292
- Shahlaei, A.; S. Torabi and M. Khosroshahli (2014). Efficiency of SCoT and ISSR markers in assessment of tomato (*Lycopersicon esculentum Mill.*) genetic diversity. *International Journal of Biosciences*, 5(2): 14-22.
- Steel, R.G.D. and J.H. Torrie (1960). *Principles and Procedures of Statistics with Special Reference to the Biological Sciences*. McGraw Hill, New York, 187-287
- Xiong, F.; R. Zhong; Z. Han; J. Jiang; L. He; W. Zhuang and R. Tang (2011). Start codon targeted polymorphism for evaluation of functional genetic variation and relationships in cultivated peanut (*Arachis hypogaea L.*) genotypes. *Mol. Biol. Rep.* 38(5): 3487-3494.

## تقدير التنوع الوراثى باستخدام واسمات SCOT وبعض الصفات المورفولوجية فى عشرة سلالات من الشعير

رحاب محمد محمد حبيبه<sup>١</sup>، جميلة عبد الحفيظ بشاشة<sup>٢</sup>، سعاد حسن حافظ<sup>٣</sup> و أحمد عبد الرحيم عبد الرحيم ليلة<sup>٣</sup>

<sup>١</sup>قسم الوراثة - كلية الزراعة - جامعة المنصورة

<sup>٢</sup>قسم البساتين - كلية الزراعة - جامعة بنى غازى

<sup>٣</sup>قسم المحاصيل - كلية الزراعة - جامعة المنصورة

### الملخص

لتقدير التنوع الجزيئى والمظهرى لعشرة سلالات من الشعير (*Hordeum vulgare* L.)، تم استخدام عشرة بادئات من SCOT بالإضافة لتقدير ١٢ صفة مورفولوجية فى موسمين (٢٠١٩/٢٠١٨ و ٢٠٢٠/٢٠١٩). هذه البادئات نجحت فى انتاج العديد من تتابعات DNA المتباينة. أظهر تكنيك SCoT تنوع جزيئى يتراوح من ٦٦,٦٧ الى ١٠٠%، وكانت قيمة متوسط قوة التحليل Rp تتراوح من ٤ الى ١١,٤٠. بالإضافة الى ان السلالات العشر اظهرت ٤١ واسمة جزيئية متنوعة ومنفردة (٢٢ موجبة و ١٩ سالبة)، اعطت السلالة B<sub>6</sub> اعلى عدد من الواسمات الجزيئية الموجبة.

تبعاً للتقييم المظهرى، كان التباين الراجع للتراكيب الوراثية عالى المعنوية لكل الصفات المدروسة وكانت قيمة معامل التوريث فى المدى الواسع تتراوح من ٤٠,٦٣ الى ٩٩,٢٢%. اعطت السلالة B<sub>7</sub> قيمة مرغوبة ومختلفة عن باقى السلالات فى أربع صفات NT/P, NS/P, (NG/S and GY/P g)، بينما اعطت كل سلالة من السلالات الأخرى قيم مرغوبة فى صفة او صفتين فقط، ولهذا فان هذه الصفات التى تميزت بها العشر سلالات من الشعير ربما يمكن ربطها مع الواسمات الجزيئية المنفردة التى تم الحصول عليها. السلالة B<sub>6</sub> أظهرت اعلى عدد من الواسمات الجزيئية الموجبة والمنفردة (سته واسمات ايجابية) فإن واحدة او اكثر من هذه الواسمات قد تكون مرتبطة مع الصفة التى أعطت فيها هذه السلالة قيم مرغوبة (GFP day) عن باقى السلالات. وهذا يدل على أن هذه الواسمات الجزيئية المنفردة ربما يمكن استخدامها كعلامات انتخابية فى برامج التربية للتحسين الوراثى لهذه الصفات فى الشعير.